

# Detection of KMT2A-PTDs in healthy donor and myeloid malignant samples using next generation sequencing

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## INTRODUCTION

*KMT2A* (*MLL*) fusions and *KMT2A*-PTD (partial tandem duplication) are vital biomarkers in myeloid malignancies traditionally detected by RT-qPCR (quantitative real-time PCR). This study utilizes next generation sequencing (NGS) with OncoPrint™ Myeloid Assay GX v2 to report the detection of *KMT2A*-PTDs in both healthy donors and myeloid malignancy samples. *KMT2A* fusions in myeloid malignant samples are also reported.

Fig 1. Estimated new cases (%) of Leukemia, Lymphoma and Myeloma in USA, 2021

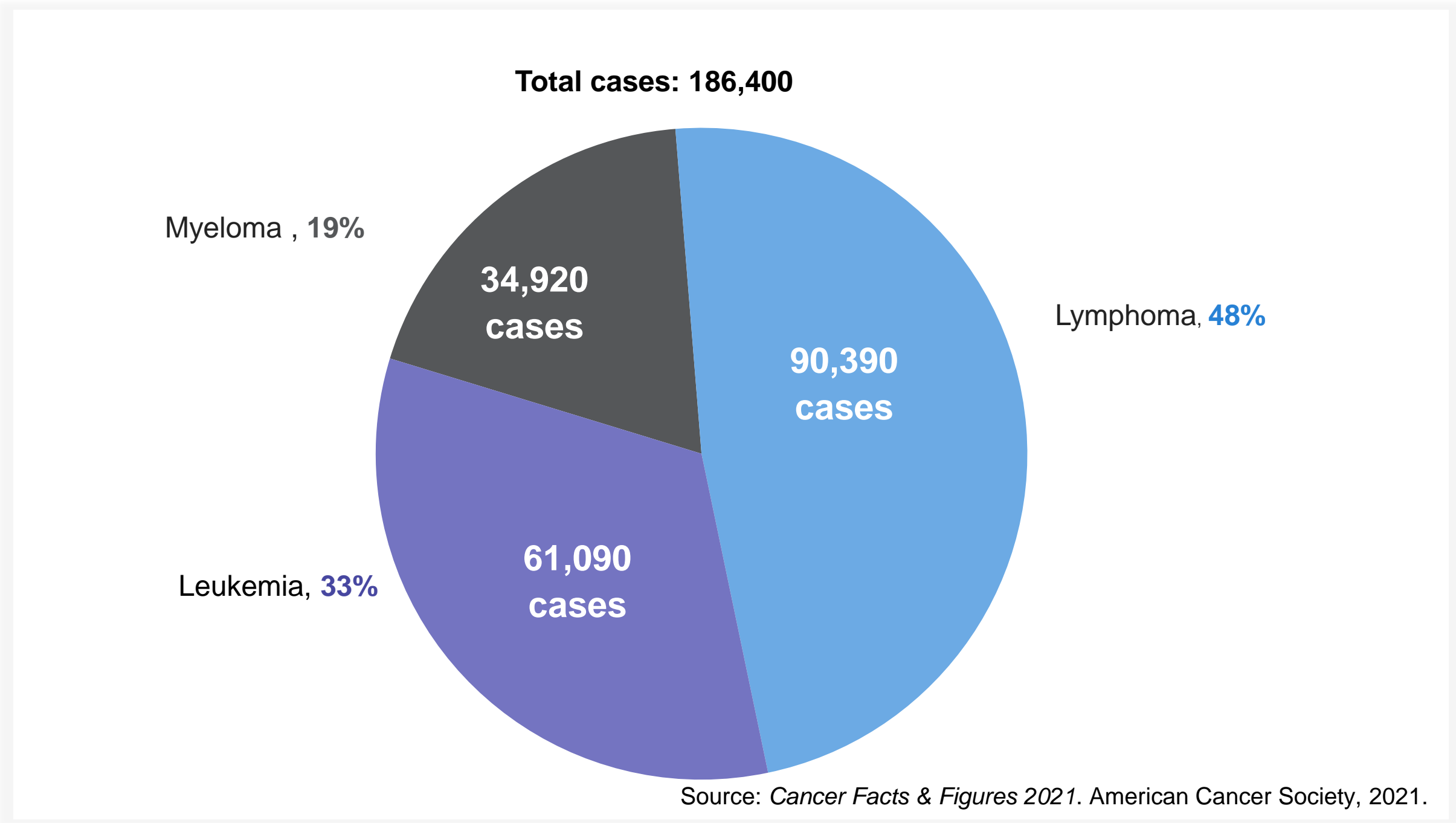


Fig 2. OncoPrint™ Myeloid assay enables rapid and efficient multi-biomarker testing

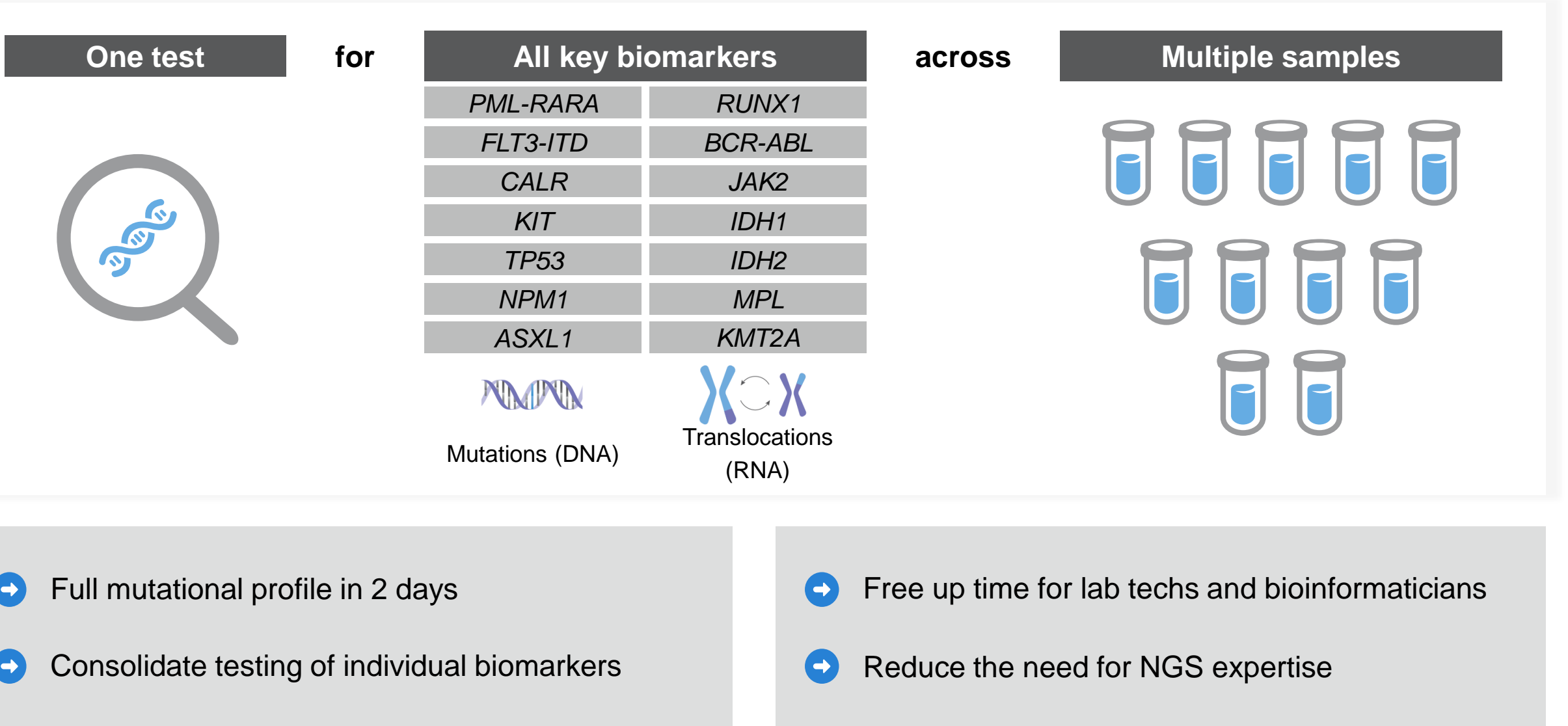


Fig 3. Genexus Instruments



## MATERIALS AND METHODS

We sequenced 8483 research samples with known myeloid malignancies in both Sonora Quest Laboratories™ and at Thermo Fisher Scientific™ South San Francisco site. We also acquired 20 healthy donor whole blood samples (total 127 replicates) from Stanford™ Blood Center and Discovery Life Sciences™ and sequenced them at 3 different sites of Thermo Fisher Scientific™ (South San Francisco, CA; Guilford, CT; Carlsbad, CA). Samples were processed on the Ion Torrent™ Genexus™ Software 6.6 and analyzed using the OncoPrint™ Myeloid Assay GX v2 for fusion profiling targeting 6 different *KMT2A*-PTD variants and 199 *KMT2A* fusion isoforms.

Table 1. OncoPrint™ Myeloid Assay GX v2 Panel

DNA Panel				RNA Panel		
Hotspot genes (28)	Full genes (17)	Fusion Driver Genes (30)	Expression genes (5)	Expression control genes (5)		
ANKRD26 ABL1 BRAF CBL CSF3R DDX41 DNMT3A FLT3 (ITD) KIT KRAS MPL MYD88 NPM1 NRAS PTPN11 SMC1A SMC3 SETBP1 SF3B1 SRSF2 IDH1 IDH2 U2AF1 JAK2 KIT	ASXL1 PRPF8 BCOR RB1 CALR RUNX1 CEBPA SH2B3 ETV6 STAG2 TET2 IKZF1 NF1 ZRSR2 PHF6	ABL1 MECOM ABL2 MET BCL2 MLLT10 BRAF MRTFA CCND1 (MKL1) CREBBP MYBL1 EGFR ETV6 FGFR1 NTRK2 FGFR2 FUS NTRK3 HMGA2 NUP214 JAK2 NUP98 KAT6A PAX5 (MOZ) PDGFRA KAT6B PDGFRB KMT2A (MLL) RARA RUNX1 KMT2A-PTDs (MLL-PTDs) TCF3 TFE3 ZNF384	BAALC MECOM MYC SMC1A WT1	EIF2B1 FBXW2 PSMB2 PUM1 TRIM27		

Fig 4. *KMT2A*-PTDs: a relevant biomarker in myeloid malignancies

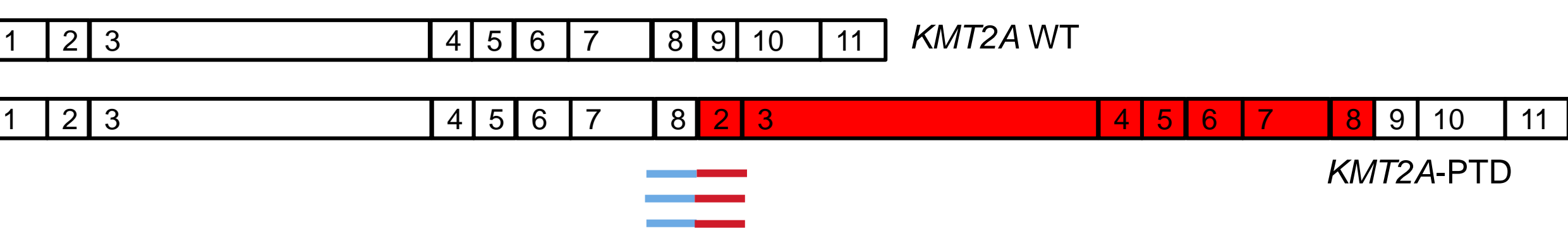


Table 2. OncoPrint™ Myeloid assay is designed to detect six *KMT2A*-PTD variants

Variant	Length (bp)
<i>KMT2A-KMT2A</i> .nt51.K11K2	178
<i>KMT2A-KMT2A</i> .K9K2	172
<i>KMT2A-KMT2A</i> .K7K2	169
<i>KMT2A-KMT2A</i> .K11K2	116
<i>KMT2A-KMT2A</i> .K8K2	105
<i>KMT2A-KMT2A</i> .K10K2	105

Table 3. Report only the PTDs associated with myeloid malignancy with high confidence

Sample name	Isoforms detected	Read count
Healthy donor sample #A	<i>KMT2A-KMT2A</i> .K8K2	9
	<i>KMT2A-KMT2A</i> .K10K2	147
	<i>KMT2A-KMT2A</i> .K7K2	201
	<i>KMT2A-KMT2A</i> .K9K2	343
Myeloid malignant sample #B	<i>KMT2A-KMT2A</i> .K10K2	551
	<i>KMT2A-KMT2A</i> .K7K2	722
	<i>KMT2A-KMT2A</i> .K8K2	311
	<i>KMT2A-KMT2A</i> .K9K2	2466

Is there a differentiation in PTD read count in myeloid malignant vs healthy donor samples?

## RESULTS

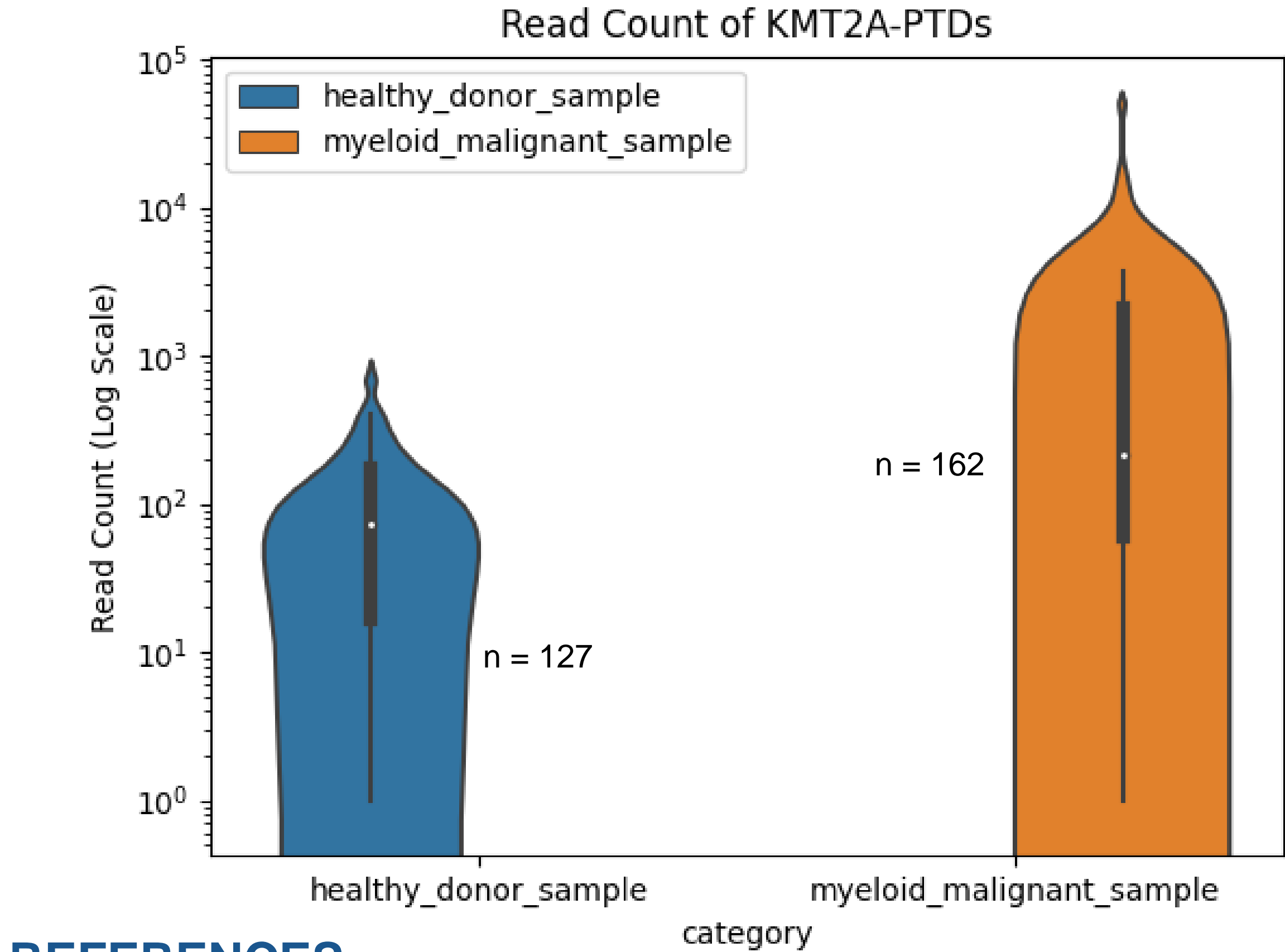
The mean read length of this data set is 90 – 120 bp and the mapped fusion reads is 20,000 – 30,000. *KMT2A*-PTDs were detected in both healthy donors and myeloid samples. Healthy donor PTD read counts were consistently <2000 and averaged 1/3 of myeloid samples. About 33% of myeloid samples had higher PTD read counts than any healthy donor sample. BLAT (BLAST™-like Alignment Tool) analysis confirmed specific exon matching on the *KMT2A* gene in both cohorts. Among the 8503 myeloid samples, 162 contained a total of 5 unique *KMT2A* PTDs, and 105 contained a total of 30 unique *KMT2A* fusion isoforms with *KMT2A-MLLT1* and *KMT2A-MLLT3* being the most prevalent *KMT2A* fusion gene pairs.

Table 4. *KMT2A* fusions & PTDs existed in ~3% samples

	Gene Pair	Found in # of unique samples	Found in % of all samples (N=8503)
<i>KMT2A</i> fusions	<i>KMT2A-AFF4</i>	2	0.02%
	<i>KMT2A-CASC5</i>	2	0.02%
	<i>KMT2A-CBL</i>	4	0.05%
	<i>KMT2A-ELL</i>	11	0.13%
	<i>KMT2A-EPS15</i>	4	0.05%
	<i>KMT2A-MLLT1</i>	30	0.35%
	<i>KMT2A-MLLT10</i>	9	0.11%
	<i>KMT2A-MLLT3</i>	29	0.34%
<i>KMT2A</i> -PTDs	<i>KMT2A-KMT2A</i>	162	1.91%
	Total	162	1.91%
Samples w/ ≥ 1 <i>KMT2A</i> fusion or PTD	Total	265	3.12%

~2% samples had at least one *KMT2A*-PTD. ~1% samples had at least one *KMT2A* fusion.

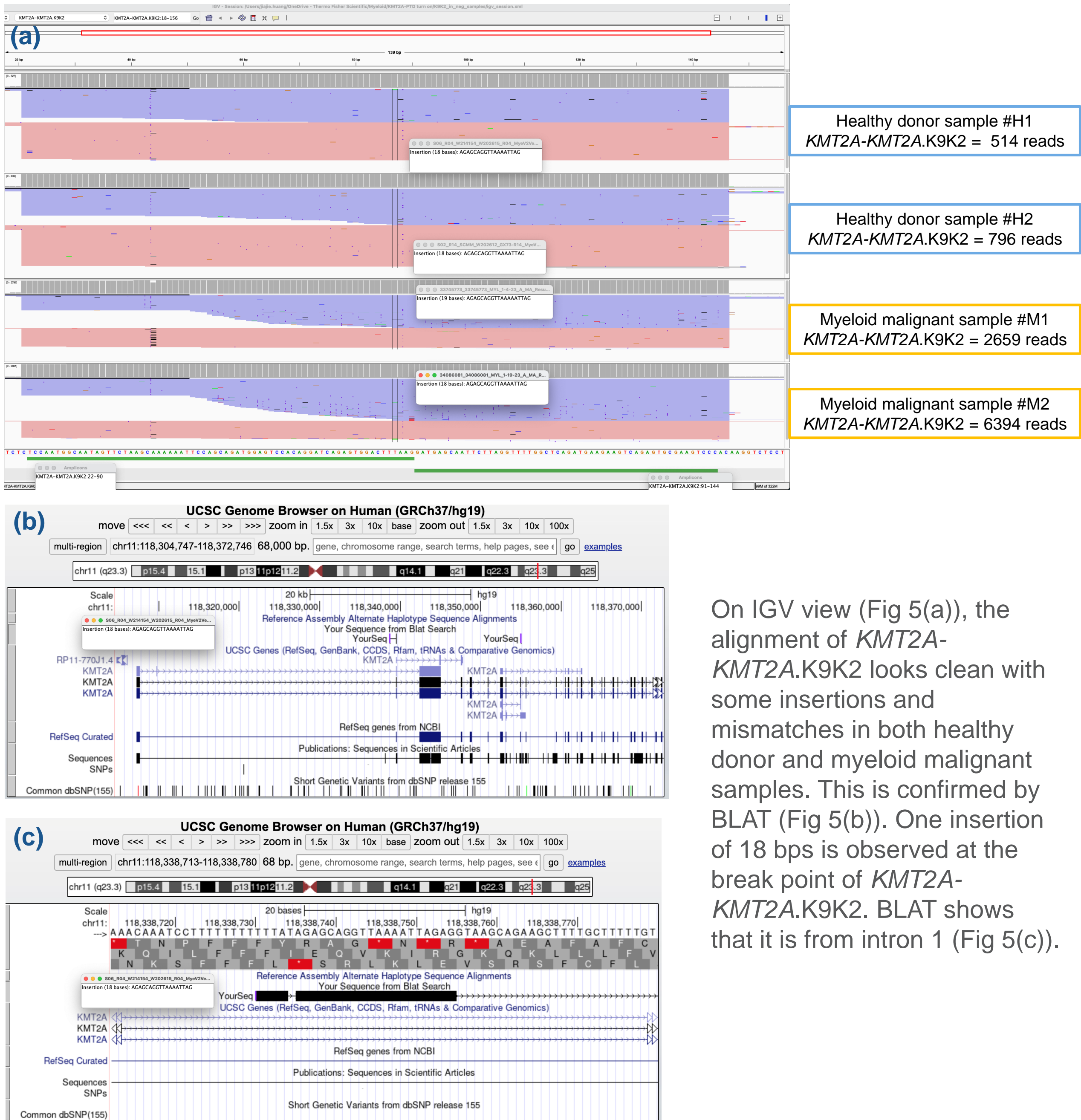
Fig 5. Read count differentiates myeloid cancer PTDs



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Dai B, Yu H, Ma T, Lei Y, Wang J, Zhang Y, Lu J, Yan H, Jiang L, Chen B. The Application of Targeted RNA Sequencing for *KMT2A*-Partial Tandem Duplication Identification and Integrated Analysis of Molecular Characterization in Acute Myeloid Leukemia. J Mol Diagn. 2021 Nov;23(11):1478-1490. doi: 10.1016/j.jmoldx.2021.07.019. Epub 2021 Aug 10. PMID: 34384895.

Fig 5. IGV view of *KMT2A-KMT2A.K9K2* in healthy donor samples vs myeloid samples



On IGV view (Fig 5(a)), the alignment of *KMT2A-KMT2A.K9K2* looks clean with some insertions and mismatches in both healthy donor and myeloid malignant samples. This is confirmed by BLAT (Fig 5(b)). One insertion of 18 bps is observed at the break point of *KMT2A-KMT2A.K9K2*. BLAT shows that it is from intron 1 (Fig 5(c)).

## CONCLUSIONS

In this study, we describe the detection of *KMT2A* fusions in myeloid malignant samples. Our study also describes the detection of *KMT2A*-PTDs in both healthy donor and myeloid samples, with myeloid cases showing significantly higher PTD read counts. additional studies to understand the relevant expression level of PTD are in progress. This intriguing finding opens opportunities for prospective studies to monitor individuals with elevated PTD levels for myeloid malignancy development and retrospective studies to explore whether healthy donors identified with this alteration years ago after blood donation were subsequently recorded in the national health system with myeloid malignancies.

## ACKNOWLEDGEMENTS

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