

Connecting patients everywhere to precision oncology

Oncomine Dx Express Test (CE-IVD)



The OncoPrint Dx Express Test—a true end-to-end solution, from specimen to clinical report

The Ion Torrent™ OncoPrint™ Dx Express Test enables laboratories to deliver clinically relevant genomic profiling in as little as 24 hours to aid clinicians in making timely therapy decisions.

This automated, true end-to-end solution—from a single supplier, with as little as 20 minutes of hands-on time—can be

implemented in a broad spectrum of clinical labs, even without next-generation sequencing (NGS) experience. The Ion Torrent™ OncoPrint™ Reporter Dx software provides biomarker results matched to approved therapies, guidelines, clinical trials, and peer-reviewed literature to aid clinicians in therapy management of cancer patients.

The OncoPrint Dx Express Test enables:



Guideline recommendations—Content covers gene targets recommended by professional guidelines for multiple solid tumors including substitutions, insertions and deletions (indels), copy number variants (CNVs), and fusions and splicing variants across 46 genes, such as *EGFR*, *BRAF*, *KRAS*, *ERBB2*, *MET*, *ALK*, *ROS1*, *RET*, and *NTRK1/2/3*, among others (Table 1).



Efficient use of samples—Requiring only 10 ng of DNA and 10 ng of RNA extracted from as little as two 5-micron formalin-fixed, paraffin-embedded (FFPE) slides, results can be generated from limited tissue and small biopsies. Plasma from liquid biopsy provides an additional sample type.



Fast results—Results can be generated in as little as 24 hours, enabling integration of molecular and immunohistochemistry (IHC) results into one complete report to aid clinicians in making timely therapy decisions.

Example Clinical Lab		Example Labs 123 Street City, State 00000 Tel +1 555-000-0000 www.example.com	
Case Number:	1234	Date:	10 Apr 2022
Sample Name:	1234	Primary Tumor Site:	Cervical Cancer
Sample Type:	Uterine Cervix		
Gender:	Unknown		
Sample Cancer Type: Non-Small Cell Lung Cancer			
Relevant Non-Small Cell Lung Cancer Findings			
Gene	Finding	Gene	Finding
ALK	ALK Inbalance	NTRK3	None detected
BRAF	BRAF V600E	NTRK2	None detected
EGFR	None detected	NTRK1	None detected
ERBB2	None detected	RET	None detected
KRAS	None detected	ROS1	None detected
MET	None detected		
Relevant Biomarkers			
Relevant Biomarker	Relevant Therapy (in this cancer type)	Relevant Therapy (in other cancer types)	
BRAF V600E ALK Frequency 63.1%	dabrafenib + trametinib	lorlatinib + anastrozole dabrafenib + MEK inhibitor dabrafenib + trametinib dabrafenib + MEK inhibitor dabrafenib + trametinib	
Public data sources included in relevant therapies (1/2021)			
Prevalent cancer biomarkers without relevant evidence based on included data sources			
ALK Inbalance			
Biomarker Descriptions			
ALK (ALK receptor tyrosine kinase)			
Background: The ALK gene encodes the ALK receptor tyrosine kinase (RTK) with sequence similarity to the insulin receptor subfamily 2 (INSR). ALK is the target of recurrent alterations in cancer, the most common being chromosomal rearrangements that generate fusion genes containing the intact ALK tyrosine kinase domain combined with multiple partner genes. ALK fusion kinases are constitutively activated and drive oncogenic transformation via activation of downstream STAT3, PI3K/AKT/mTOR, and RAS/MAPK/MEK/ERK pathways (1).			
Alterations and prevalence: ALK was discovered by positional cloning of translocations involving nucleophosmin (NPM) on 5q35 with a previously unidentified 5'K on 2Q22 (NALK), which occur in over 50% of anaplastic large cell lymphoma cases (2). In contrast, about 5% of non-small cell lung cancer (NSCLC) cases generate recurrent ALK fusions with EML4, KIF5B, and HPIV1 (3).			
Optional left-aligned text block, one line on multiple lines. Lines will span full width of page width and then wrap to new line within footer area.			
Disclaimer: The data presented here is for informational purposes only and does not constitute a recommendation. The data version is 2021-04-01. The content of this report may be subject to change without notice.			

OncoPrint Reporter Dx software

OncoPrint Reporter Dx software matches genomic variant information with relevant therapies, guidelines, clinical trials, and peer-reviewed literature. It is an intuitive software that produces a clear and concise biomarker report without requiring specialized bioinformatics experience.

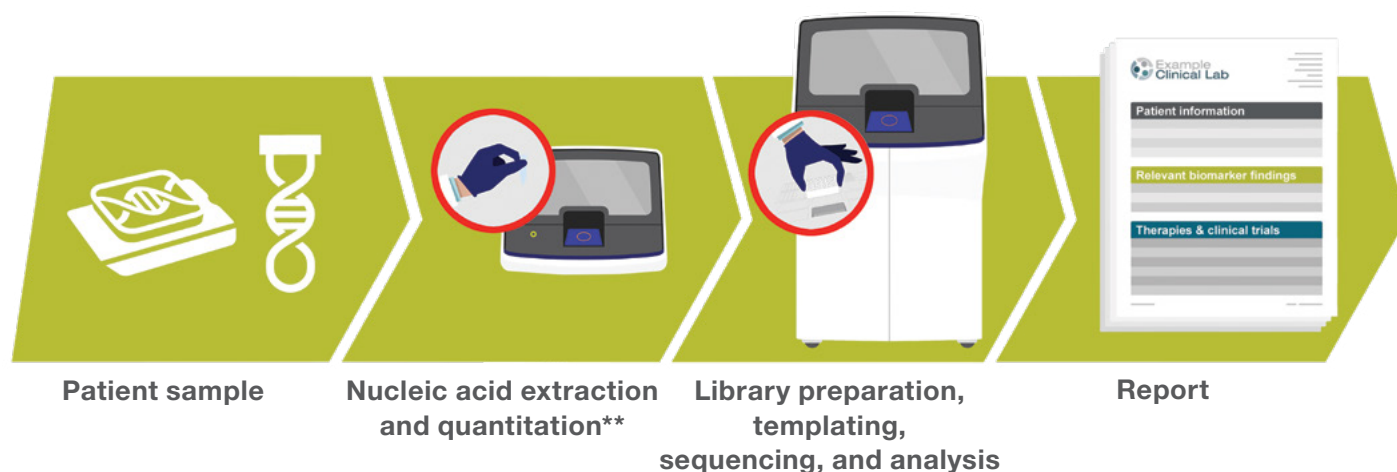
A true end-to-end solution from one supplier

The Ion Torrent™ Genexus™ Dx System automates the NGS workflow, from the patient sample to report, and delivers results in as little as 24 hours with just two user touchpoints.*

With automated library preparation, sequencing, and analysis involving as little as 20 minutes of hands-on time, the Oncomine Dx Express Test on the Genexus Dx System helps reduce laboratory staff burden and the potential for human errors.

The Genexus Dx software facilitates tracking sample information through the workflow. On-instrument analysis and local reporting alleviate the need for specialized bioinformatics experience.

* Timing varies by number of samples and sample type.



** The Genexus Dx Purification System will be available at a later date.

Table 1. Oncomine Dx Express Test gene list.

Deletions, insertions, and substitutions				Copy number alterations	Fusions and splicing variants	
<i>AKT1</i>	<i>ERBB2</i>	<i>IDH1</i>	<i>NTRK2</i>	AR	<i>ALK</i>	<i>NTRK1</i>
<i>AKT2</i>	<i>ERBB3</i>	<i>IDH2</i>	<i>NTRK3</i>	EGFR	AR	<i>NTRK2</i>
<i>AKT3</i>	<i>ERBB4</i>	<i>KEAP1</i>	<i>PDGFRA</i>	ERBB2	BRAF	<i>NTRK3</i>
<i>ALK</i>	<i>ESR1</i>	<i>KIT</i>	<i>PIK3CA</i>	ERBB3	EGFR	NUTM1
<i>AR</i>	<i>FGFR1</i>	<i>KRAS</i>	<i>PTEN</i>	FGFR1	ESR1	<i>RET</i>
<i>ARAF</i>	<i>FGFR2</i>	<i>MAP2K1</i>	<i>RAF1</i>	FGFR2	FGFR1	<i>ROS1</i>
<i>BRAF</i>	<i>FGFR3</i>	<i>MAP2K2</i>	<i>RET</i>	FGFR3	FGFR2	RSPO2
<i>CDK4</i>	<i>FGFR4</i>	<i>MET</i>	<i>ROS1</i>	KRAS	FGFR3	RSPO3
<i>CHEK2</i>	<i>FLT3</i>	<i>NRAS</i>	<i>STK11</i>	MET	<i>MET</i>	
<i>CTNNB1</i>	<i>GNAS</i>	<i>NTRK1</i>	<i>TP53</i>	PIK3CA	NRG1	
<i>EGFR</i>	<i>HRAS</i>					

Genes in bold are only available for FFPE samples.

OncoPrint Dx Express Test performance—FFPE samples

Extensive performance studies were conducted to establish performance characteristics of the OncoPrint Dx Express Test for FFPE samples. For complete studies and results, refer to the OncoPrint Dx Express Test User Guide.

Analytical accuracy study

The analytical accuracy was evaluated with 151 clinical FFPE samples from 6 cancer types (breast cancer, colorectal cancer (CRC), glioma, melanoma, non-small cell lung cancer (NSCLC), and thyroid cancer). The variants evaluated included single-nucleotide variants (SNVs), indels, CNVs, and fusions (Table 2). The concordance evaluation study included:

- 75 variant-negative and 76 variant-positive specimens
- 2 sites
- 2 NGS-based orthogonal reference assays: Reference Assay 1 and Reference Assay 2

The positive percent agreement (PPA) and negative percent agreement (NPA) were defined as the proportion of variant-positive and variant-negative specimens, respectively, as determined by the reference methods that were also determined by the OncoPrint Dx Express Test.

Analytical accuracy results are summarized in Table 3.

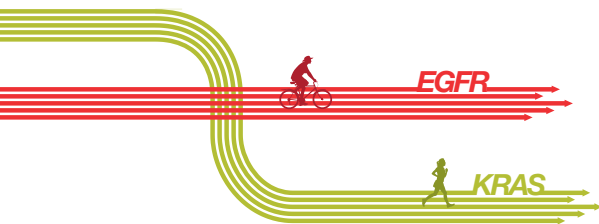


Table 2. Variant description for FFPE sample.

Indication	Gene	Variant	Variant type
Breast	<i>PIK3CA</i>	E545 or H1047	SNV
	<i>PIK3CA</i>	Amplification	CNV
	<i>ERBB2</i>	Amplification	CNV
	<i>NTRK</i>	<i>NTRK3</i>	Fusion
CRC	<i>KRAS</i>	G12C	SNV
	<i>BRAF</i>	V600K or V600E	SNV
	<i>NTRK</i>	<i>NTRK1</i>	Fusion
	<i>EGFR</i>	L858R	SNV
NSCLC	<i>EGFR</i>	T790M	SNV
	<i>BRAF</i>	V600E	SNV
	<i>KRAS</i>	G12C	SNV
	<i>EGFR</i>	Exon 19 deletion	Indel
	<i>EGFR</i>	Exon 20 insertion	Indel
	<i>ERBB2</i>	Exon 20 insertion	Indel
	<i>ALK</i>	Fusion	Fusion
	<i>ROS1</i>	Fusion	Fusion
Melanoma	<i>RET</i>	Fusion	Fusion
	<i>MET</i>	MET exon 14 skipping	Alternative splice form
	<i>MET</i>	Amplification	CNV
	<i>BRAF</i>	V600K or V600E	SNV
	<i>NTRK</i>	<i>NTRK1</i>	Fusion
Thyroid	<i>BRAF</i>	V600K or V600E	SNV
	<i>RET</i>	Mutations	SNV
	<i>RET</i>	Fusion	Fusion
Glioma	<i>NTRK</i>	<i>NTRK1</i> and <i>NTRK3</i>	Fusion
	<i>IDH1</i>	R132	SNV
	<i>IDH2</i>	R172	SNV

Table 3. OncoPrint Dx Express Test—concordance.

	Variant type	Reference assay	Percent agreement (%)	95% CI
PPA	SNVs and indels	1	93.44 (57/61)	(84.05, 98.18)
NPA	SNVs and indels	1	99.99 (43,026/43,029)	(99.98, 100.00)
PPA	CNVs	2	100.00 (27/27)	(87.23, 100.00)
NPA	CNVs	2	99.30 (283/285)	(97.49, 99.91)
PPA	Fusions	1	91.67 (11/12)	(61.52, 99.79)
NPA	Fusions	1	99.98 (11,642/11,644)	(99.94, 100.00)

Limit of blank (LOB) study

The LOB was established by profiling 30 clinical FFPE samples confirmed to be variant-negative by a reference method. The study included:

- **2 replicates per sample**
- **2 reagent lots**
- **11 tissue types: bladder, brain, breast, bile duct, colon, endometrium, lung, pancreas, prostate, skin, and thyroid**

For all 30 samples, the false-positive rate of the test was determined to be 0.75% for SNVs, 0% for indels, 0% for CNVs, and 0% for fusions. By definition of the Clinical and Laboratory Standards Institute (CLSI) EP17-A2, the LOB is zero.

Limit of detection (LOD) study

The LOD was evaluated with 20 representative SNVs, indels, CNVs, and RNA fusions detected by the Oncomine Dx Express Test in clinical FFPE samples. The LOD is defined as the lowest variant level that can be detected at least 95% of the time.

Clinical specimens representing 6 cancer types (breast cancer, CRC, glioma, NSCLC, melanoma, and thyroid cancer) were used as the source of DNA and RNA. Variant-containing specimens were blended with wild type samples, and the study included:

- **6 titration levels**
- **2 reagent lots**
- **10 replicates per sample blend**

Based on a representative approach, the LODs ranged from:

- **3.07–6.48% allelic frequencies for SNVs and indels (mean = 4.29% allelic frequency)**
- **4.91–5.32 copies for CNVs**
- **5.27–12.35 molecular counts (median = 8.85 molecular counts) and 7.87–207.5 reads for fusions**

Precision study

The repeatability and reproducibility were evaluated using 20 representative DNA variants and RNA fusions in FFPE samples from 6 cancer types: breast cancer, CRC, glioma, melanoma, NSCLC, and thyroid cancer.

Three sites, with two operators and instruments per site, were used for the study. DNA and RNA were extracted from clinical FFPE samples, then blended with wild type DNA or RNA into 7 DNA blends and 7 RNA blends. Two levels per blend were generated and distributed to sites and operators for testing.

The mean call rates excluding no-calls were 99.23%, 100%, and 99.69% for variant-positive SNVs/indels, CNVs, and fusions, respectively. The mean call rate excluding no-calls was 100% for wild type DNA (negative-calls) and wild type RNA.

For details, see the Oncomine Dx Express Test User Guide.



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Oncomine Dx Express Test performance—plasma samples

Extensive performance studies were conducted to establish performance characteristics of the Oncomine Dx Express Test for cell-free total nucleic acid (cfTNA). For complete studies and results, refer to the Oncomine Dx Express Test User Guide.

Analytical accuracy study

The analytical accuracy of the Oncomine Dx Express Test for plasma was evaluated with 80 plasma samples from NSCLC comprising 40 variant-positive and 40 variant-negative samples (Table 4). The concordance study was performed at two sites that received an identical set of samples. One site used the Oncomine Dx Express Test, and the second site used an NGS-based reference assay.

The PPA and NPA were defined as the proportion of variant-positive and variant-negative specimens, respectively, as determined by the reference method and the Oncomine Dx Express Test.

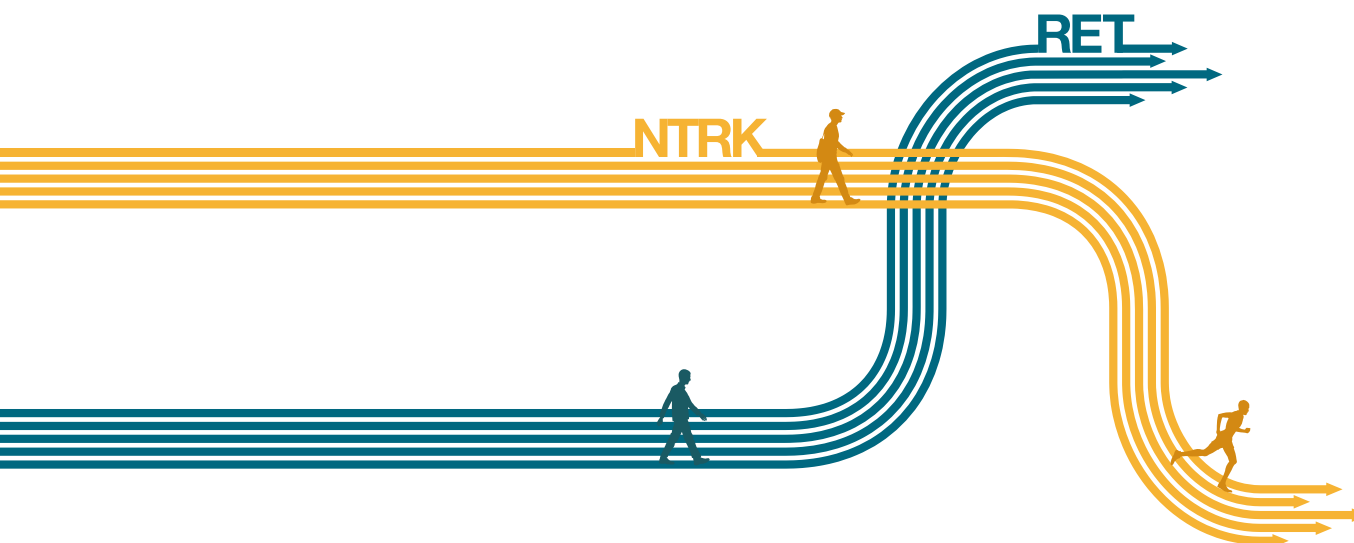
Analytical accuracy results are summarized in Table 5.

Table 5. Oncomine Dx Express Test—concordance.

	Alteration type	Percent agreement (%)	95% CI
PPA	SNVs and fusions	100.00 (51/51)	(93.02, 100.00)
NPA	SNVs and fusions	99.98 (56,272/56,282)	(99.97, 99.99)

Table 4. Variant description for plasma samples.

Gene	Variant	Variant type
<i>ERBB2</i>	SNV	SNV
<i>EGFR</i>	L858R	SNV
<i>EGFR</i>	T790M	SNV
<i>BRAF</i>	V600E	SNV
<i>KRAS</i>	G12C	SNV
<i>EGFR</i>	Exon 19 deletion	Indel
<i>EGFR</i>	Exon 20 insertion	Indel
<i>ERBB2</i>	Exon 20 insertion	Indel
<i>ALK</i>	Fusion	Fusion
<i>ROS1</i>	Fusion	Fusion
<i>RET</i>	Fusion	Fusion
<i>MET</i>	Fusion	Fusion



Limit of blank (LOB) study

The LOB was established by profiling cfTNA extracted from 30 blood plasma samples from healthy donors confirmed to be variant-negative by a reference method. The study included:

- **2 replicates per sample**
- **2 reagent lots**

For all 30 samples, the false-positive rate was determined to be 0.20% for SNVs, 0% for indels, and 0% for fusions. By definition of CLSI EP17-A2, the LOB is zero.

Limit of detection (LOD) study

The LOD was evaluated with 11 representative SNVs, indels, CNVs, and RNA fusions detected by the Oncomine Dx Express Test in clinical plasma samples. The LOD is defined as the lowest variant level that can be detected at least 95% of the time.

The study included:

- **6 titration levels**
- **2 reagent lots**
- **10 replicates per sample blend**
- **2 cfTNA input levels: 5 ng and 30 ng**

Based on a representative variant approach, the LODs for SNVs and indels ranged from 0.65% to 1.82% allelic frequency (mean = 1.9% allelic frequency) for the 5 ng input level. The LODs for SNVs and indels at the 30 ng input level ranged from 0.31% to 0.42% allelic frequency (mean = 0.36% allelic frequency).

The LODs for RNA fusions at the 5 ng input level ranged from 9.9 to 19.6 molecular counts (median = 14.3 molecular counts). The LODs for RNA fusions at the 30 ng input level ranged from 6.4 to 8.0 molecular counts (median = 7.5 molecular counts).

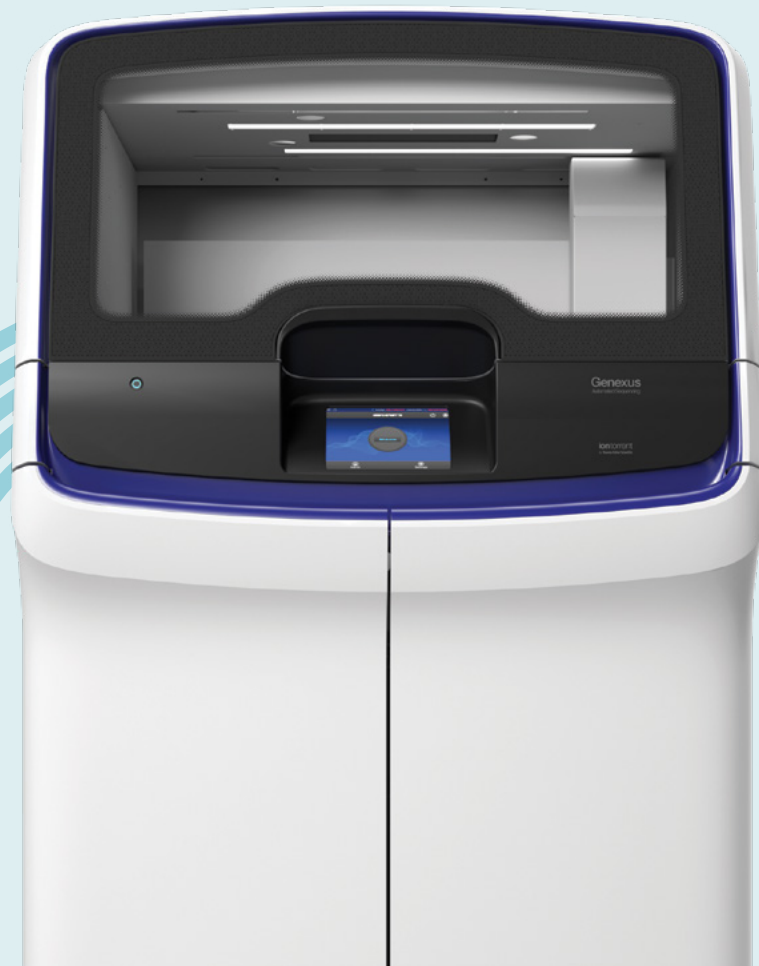
Precision study

The repeatability and reproducibility were evaluated using contrived cfTNA plasma samples prepared by blending cfTNA extracted from variant-positive cell lines and cfTNA from plasma of healthy donors.

Three sites, with two operators and instruments per site, were used for the study. Site 1 had 4 instruments, and sites 2 and 3 had 2 instruments each.

The mean call rates excluding no-calls were 99.86% and 99.25% for variant-positive SNVs/indels and fusions, respectively. The mean call rate excluding no-calls was 100% for wild type DNA (negative-calls) and wild type RNA.

For details, see the Oncomine Dx Express Test User Guide.



OncoMine Dx Express Test

The following reagents and supplies are available for order as needed. For detailed contents and storage information, see the OncoMine Dx Express Test User Guide.

Ordering information

Product	Cat. No.
Genexus Dx Integrated Sequencer	A53579
Genexus Dx Barcodes 1-32 HD	A54104
Genexus Dx Pipette Tips	A54105
Genexus Dx Library Strips 1 and 2-HD	A50430
Genexus Dx GX5 Chip and Genexus Coupler	A54106
Genexus Dx Templating Strips 3-GX5 and 4	A50431
Genexus Dx Sequencing Kit	A50432
OncoMine Dx Express Test Panel	A54103
OncoMine Dx Express Test FFPE DNA and RNA Control Kit	A52167
OncoMine Dx Express Test Plasma cfTNA Control Kit	A52168
OncoMine Reporter Dx, one-year license	A54966

 Learn more at thermofisher.com/oncomine-dxexpress

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Abbreviated Intended Use: The OncoMine Dx Express Test is a qualitative *in vitro* diagnostic test that uses targeted next-generation sequencing (NGS) technology, the Genexus Dx System, to detect deletions, insertions, substitutions, and copy number gain present in 42 genes and fusions in 18 genes from DNA and RNA extracted from FFPE tumor tissue samples. OncoMine Dx Express Test also detects deletions, insertions, and substitutions in 42 genes and fusions in 7 genes from cfTNA extracted from plasma samples. The OncoMine Dx Express Test is intended to provide clinically relevant tumor mutation profiling information to be used by qualified health care professionals in accordance with professional guidelines as an aid in therapy management of cancer patients with solid malignant neoplasms using FFPE samples and as an aid in therapy management of cancer patients with NSCLC using plasma samples. It is not conclusive or prescriptive for labeled use of any specific therapeutic product.